



July, 2012

EnSpire

Multilabel Plate Reader

博克科技有限公司
產品應用專員 曾筱筑

EnSpire Modular design with individual fixed optics

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- Fluorescence Intensity
- UV/Absorbance
- Ultra-sensitive luminescence
- Alpha technology
- High-end Temperature control (up to 65°C)
- 2 Injectors with heating and Stirrer function
- Touch Screen
- Intuitive, user friendly software

PKI's high performance
Quad monochromator technology



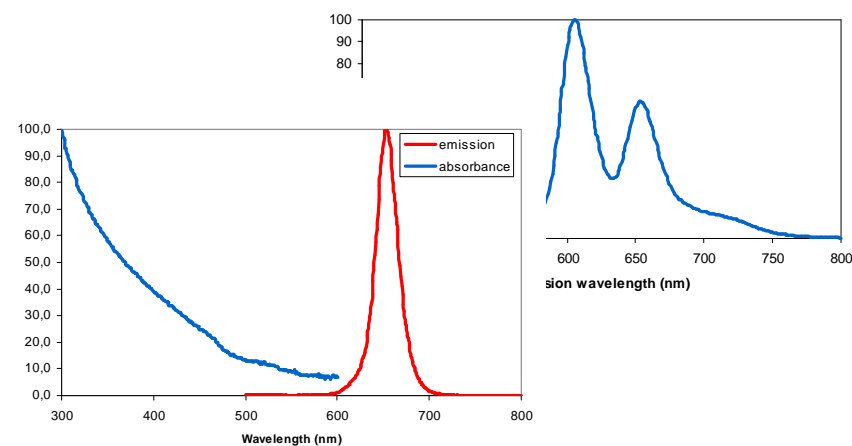
EnSpire with Quad Monochromators



EnSpire (monochromator)

FOR QUANTITATION

Absorbance & Fluorescence

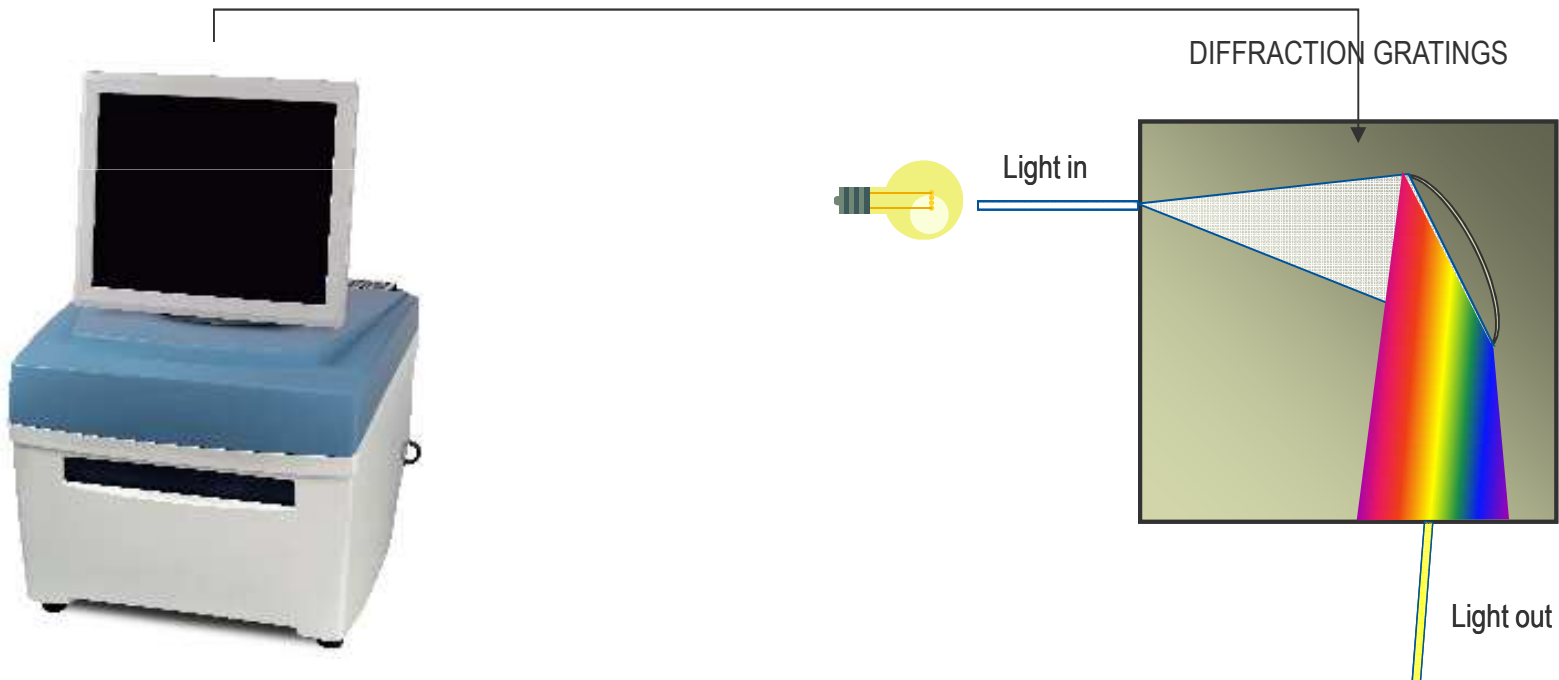


EnSpire Monochromator Applications

1. Fluorescent assays
2. Absorbance assays
 - ELISA (absorbance)
 - Quantitation assays (UV/VIS absorbance)
 - Protein (A280)
 - Bradford (A405)
 - DNA/RNA (A260/A280)



- Monochromators use diffraction gratings to physically separate the individual wavelengths present in the white light → easy wavelength selection and scanning



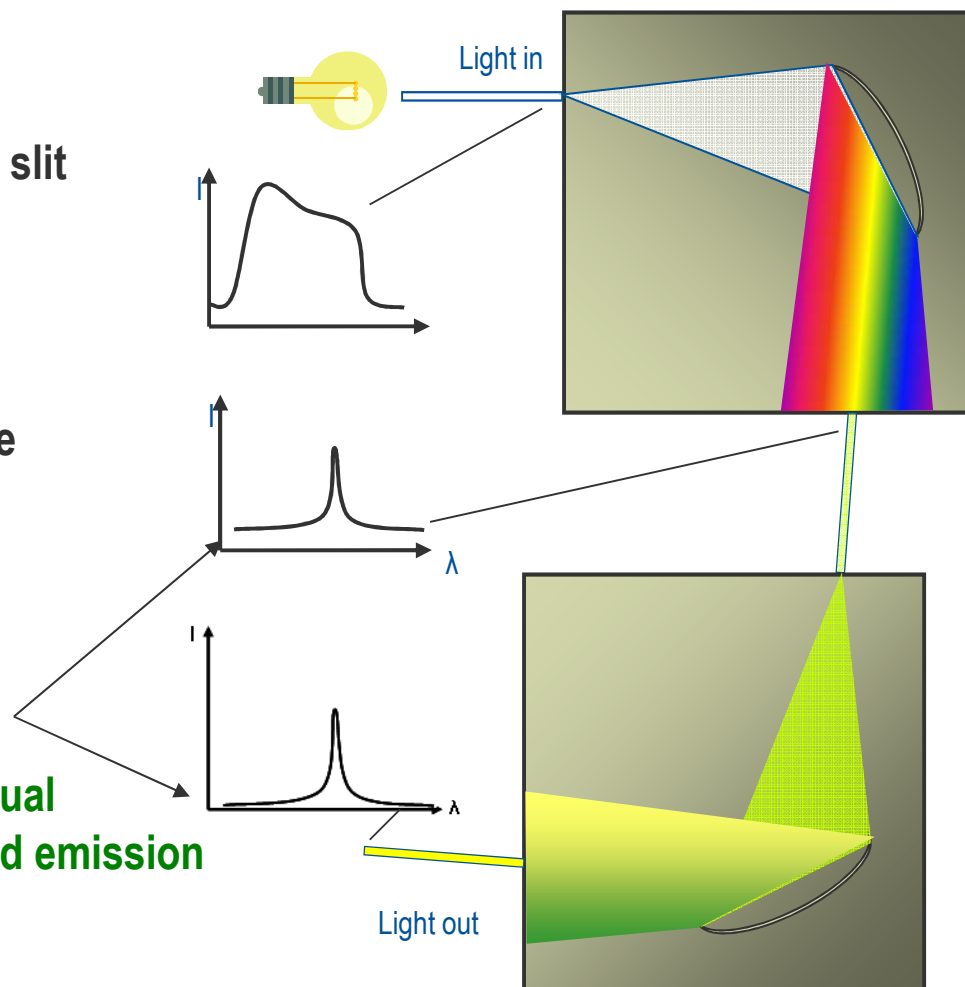
Benefits of Quad Monochromators

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- The filtering capacity of single monochromator is limited
 - Unwanted wavelengths pass the slit
 - Increased background noise

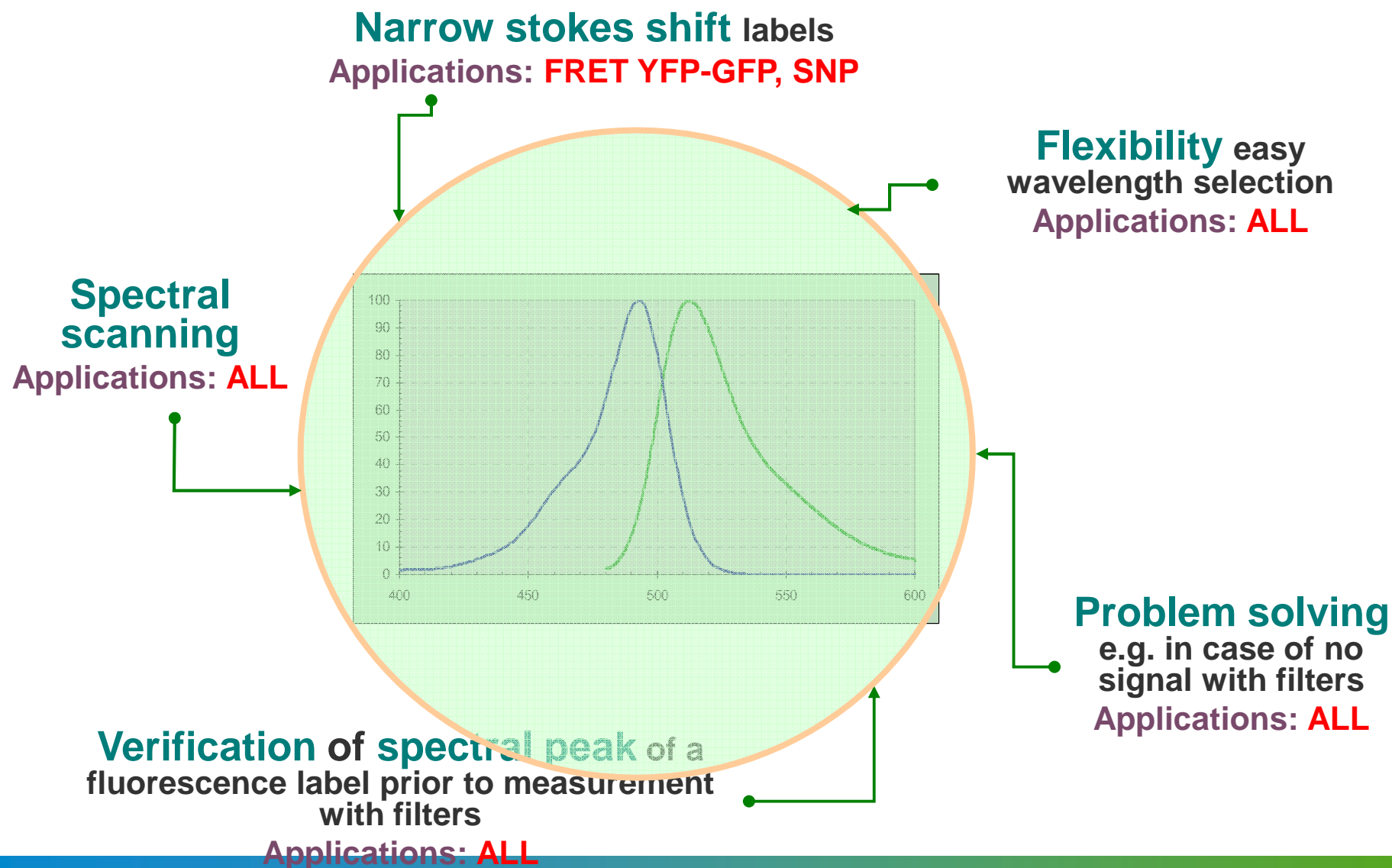
- Dual monochromator filters light twice
 - Lower assay background noise
 - Higher S/B

A Quad monochromator system uses dual monochromators for both excitation and emission



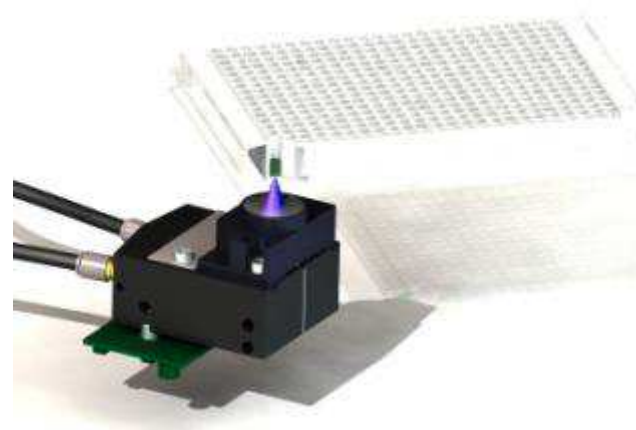
Features and benefits – Monochromators

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EnSpire with Monochromator

- Wavelength is selectable from 220 nm to 850 nm.
- Wavelength selection for excitation & emission
- Tunable in 0.1 nm increment
- Bottom-reading



EnSpire with Monochromator

- The excitation is done from top and detection is done from bottom.
- Wavelength range: 230-1000 nm

EnSpire with Alpha technology

AlphaScreen and AlphaLISA technology



AlphaLISA™

Say good-bye to all those wash steps.

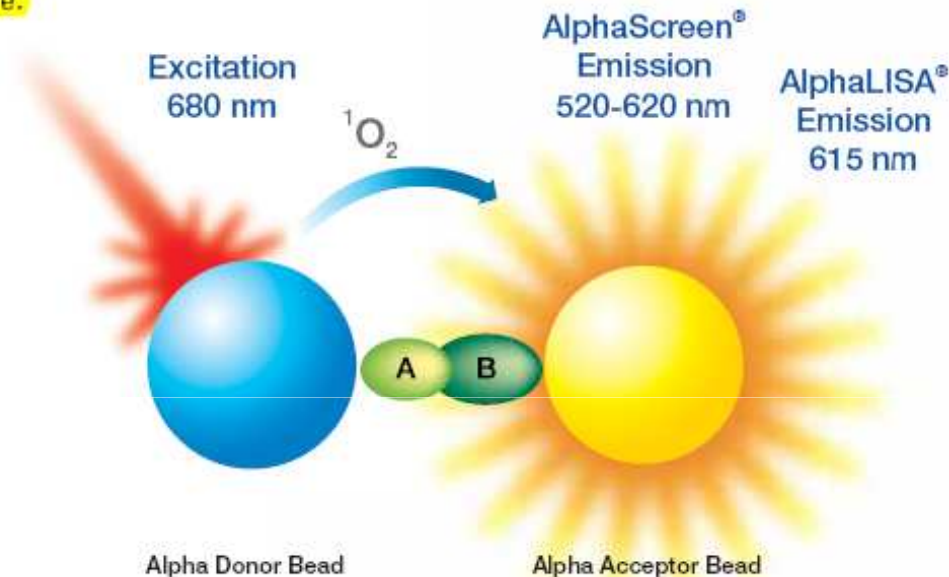
Alpha technology

Amplified luminescent proximity homogeneous assay

- Amplified → Signal intensity
- Luminescent → Reaction measurement
- Proximity → Distance between Alpha Donor and Acceptor beads
- Homogeneous → No wash necessary
- Mix and measure!

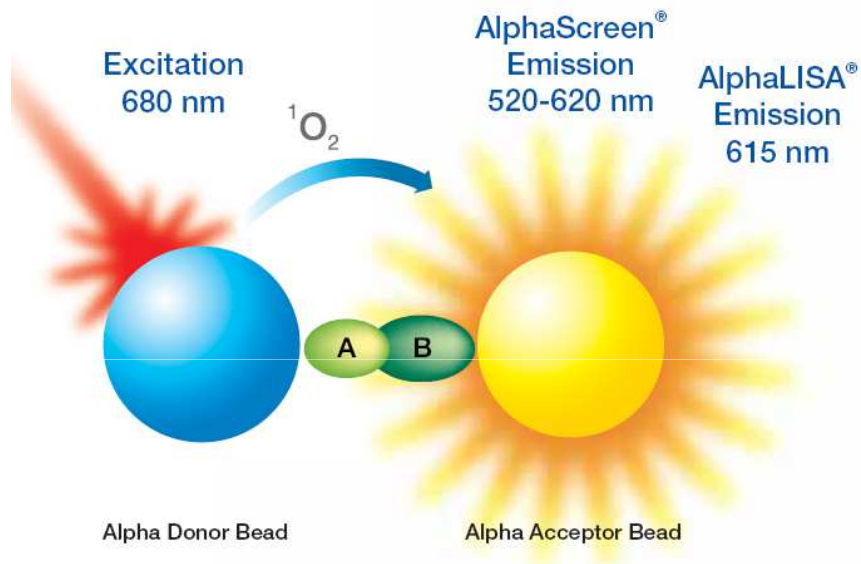
Illumination at a long wavelength ensures that few biological or assay substances will interfere.

A high concentrate of photosensitizer in each Alpha Donor bead generates up to 60,000 singlet oxygen molecules per second. This results in a very high signal amplification that contributes to detection sensitivity to the attomole level.



Singlet oxygen can travel up to 200 nm in solution, allowing the measurement of very large biological molecules.

The Acceptor beads contain a thioxene derivative that reacts with the singlet oxygen molecule to generate a chemiluminescence reaction. This energy is transferred to fluorophores within the same bead, shifting the emission to 520-620 nm in the case of the AlphaScreen beads and 615 nm with AlphaLISA beads. A half-life decay reaction of 0.3 sec allows detection in a time-resolved mode.



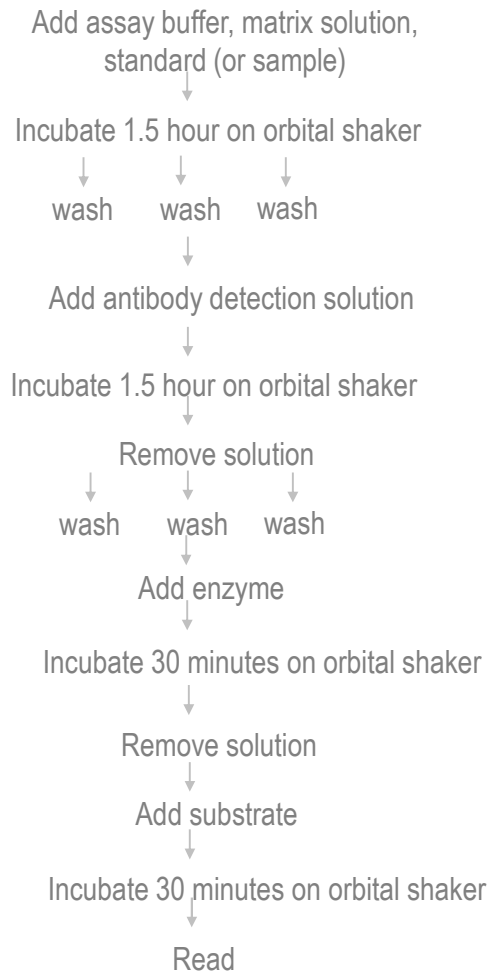
- A homogeneous, bead-based proximity platform
- Detects virtually any molecule from large endogenous protein complexes to very small peptides
- Works with a variety of sample types including, serum, plasma, cell lysates, cell supernatant and purified reagents
- NO wash steps are required, improving workflow and making it easily automated
- Available in a variety of detection kits for pre-validated assays with off-the-shelf reagents

all in one well and with no wash steps

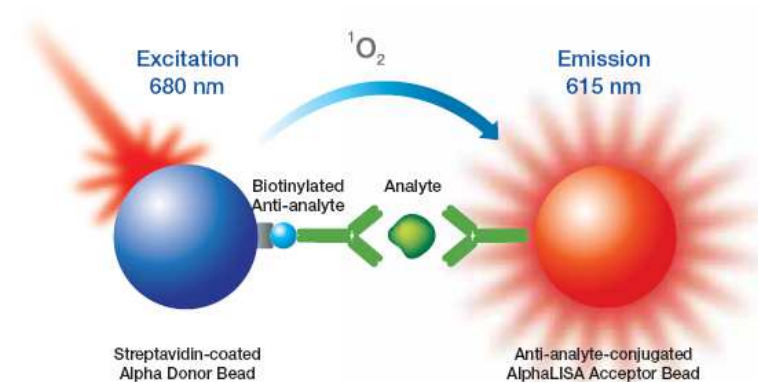
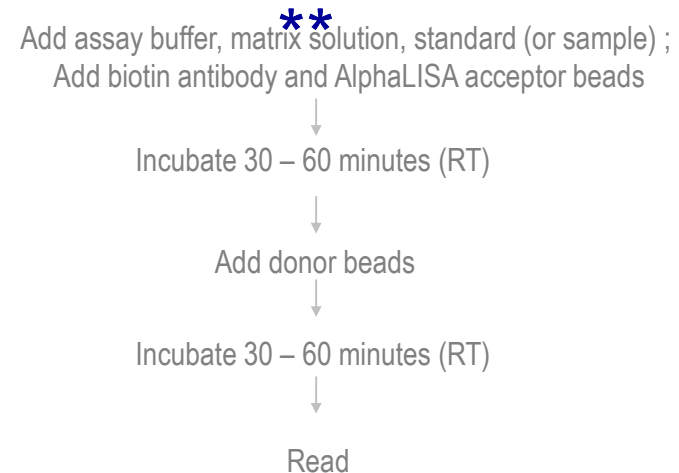
Steps of ELISA vs. AlphaLISA

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ELISA*



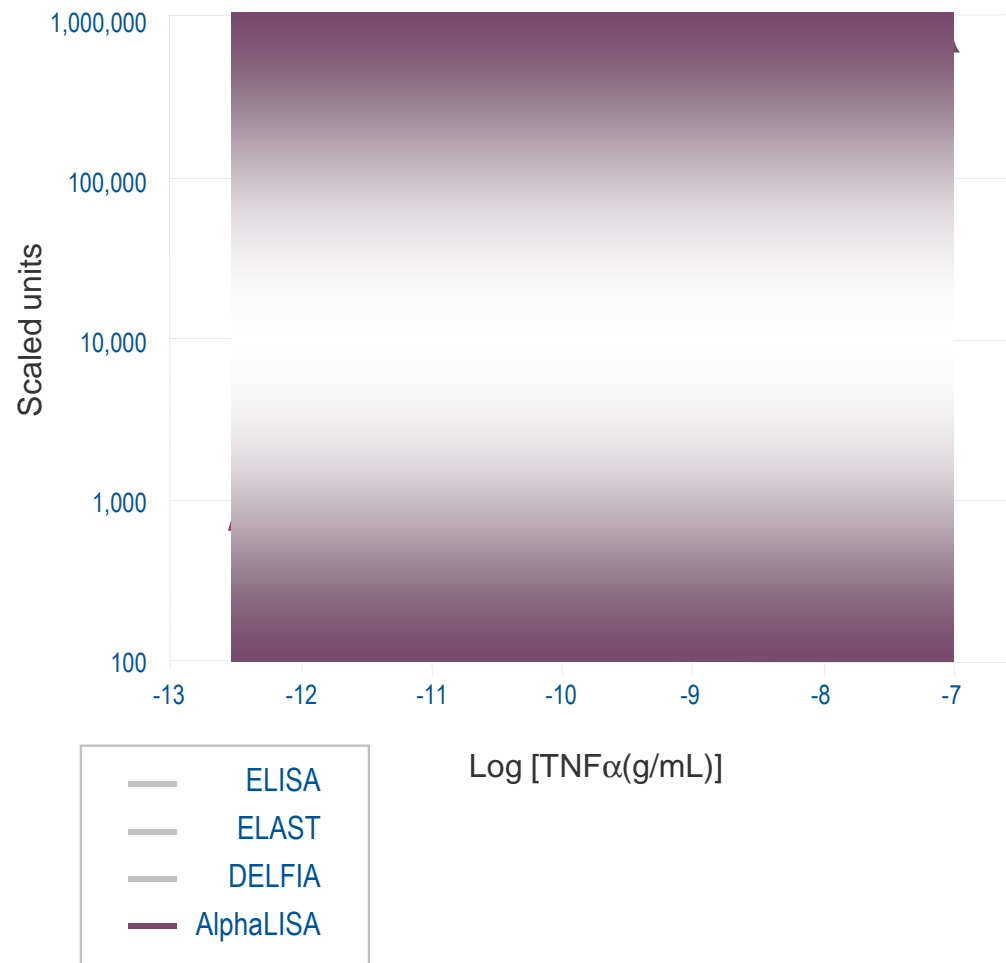
AlphaLISA



AlphaLISA Benefits

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ELISA-Related Technologies



Highly sensitive

- Save money
- Reduce need for large volume of samples

Wide dynamic range

- Decrease dilutions

Homogeneous technology

- Time for work (increased productivity)
- Time to publish
- Decrease sample prep (improve work flow)

Proximity based

- Detection of simple to large complex biological interactions
- Suited for analytes from various sources including serum and plasma

Validated on PerkinElmer Instruments

EnSpire Alpha technology

- The Alpha technology uses laser for excitation and photomultiplier tube (PMT) for detection.
- 680 nm excitation with semiconductor laser
- The photomultiplier tube is located as close as the well as possible in order to maximize the sensitivity

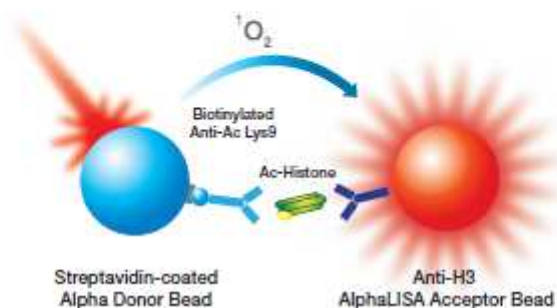
EnSpire Alpha technology

- AlphaScreen measurements are done in a “fly” mode,
 - Excitation is done for one well while the emission is done for another, during this the plate is constantly moving
- Excitation and emission collection times are adjustable in the software.



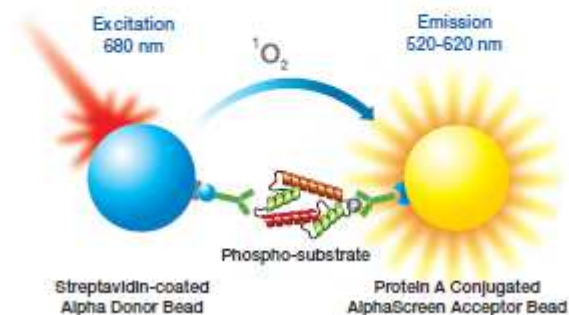
Receptor-Ligand

Acetyl-Histone Detection Assay



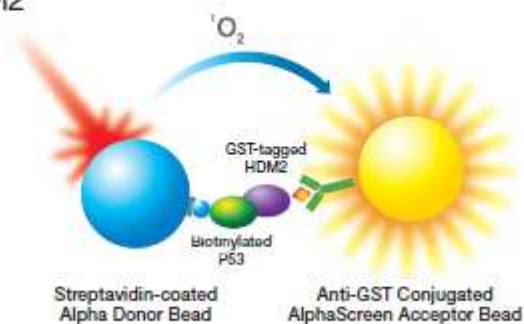
Kinase

EGFR



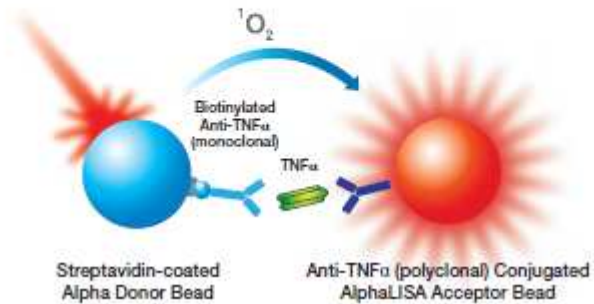
Protein:Protein

p53/HDM2



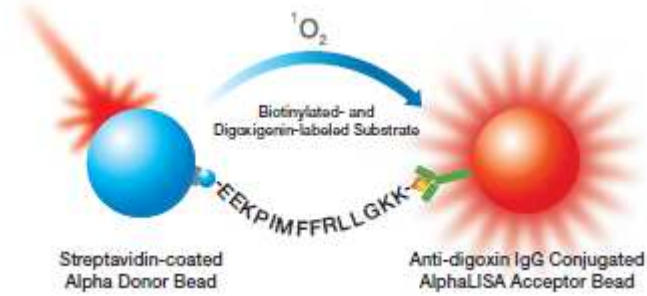
Immunoassay

TNF α detection – Sandwich format



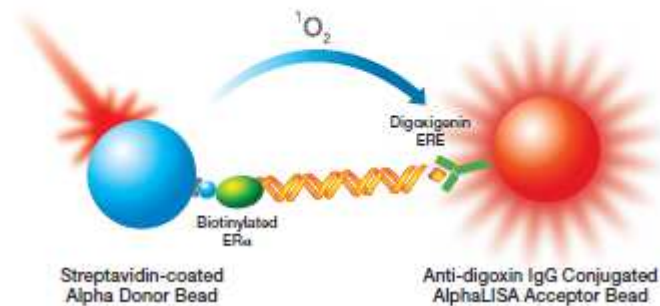
Protease

Cathepsin D – Cleavage of the specific substrate by cathepsin D results in a signal decrease



Protein-DNA Interaction

ERE-ER α interaction – Biotinylated protein



AlphaLISA® Immunoassay Kits

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Biologics	Angiogenesis	Cancer	Cardio-vascular	Inflammation	Inflammation	Neuro-degeneration
IgA	TNFα	AFP	Myeloperoxidase	CCL2 / MCP1	IFN-g (mouse)	Aβ 1-40
IgG	VEGF	CA125	NT-proBNP	CCL3/MIP-1a	IL1b (mouse)	Aβ 1-42
IgM	VEGFB	CXCL11/I-TAC	PCSK9	CCL4/MIP-1b	IL6 (mouse)	sAPPα
CHO-P	VEGFC	EGF-R	Plasminogen	COMP	IL10 (mouse)	sAPPβ
NSO-P	VEGFD	EPO	Renin/Prorenin	CRP	IL2 (mouse)	sAPPα (C-term spec.)
<i>E.Coli</i> HCP		ERBB2 / HER2	tPA	CXCL10/IP-10	IL7 (mouse)	sAPPβ (high sensitivity)
Residual Protein A	TNFα (mouse)	IFN-b	a-2 macroglobulin	G-CSF	IL15 (mouse)	Aβ 1-15/16
IgE		MMP1	ICAM-1	GM-CSF	IL17A (mouse/rat)	Aβ 1-40 (high spec.)
SerumAlbumin		MMP2		IFN-a	CCL2/MCP-1 (mouse/rat)	Aβ 1-42 (high spec.)
		MMP9		IFN-g		Tau
		b-NGF		IL1α		
		PSA		IL1β		
		TIMP1		IL2		Aβ 1-40 (mouse/rat)
		TFF3		IL3		Aβ 1-42 (mouse/rat)
		HGFR/c-MET		IL4		
		MMP3		IL5		
				IL6		
				IL7		
				IL8		
				IL10		
				IL12 (p70)		
				IL13		
				IL17		
				IL18		
				CXCL1/GRO-a		
				CXCL9/MIG		
				IL15		

Metabolic	Virology
Adiponectin	HIV p24
GH	
GLP-1	
IGF1	
IGF2	
Insulin	
Leptin	
Prolactin	
Albumin (mouse)	
C-peptide (mouse/rat)	

<http://www.perkinelmer.com/surefire>

EnSpire ultra-sensitive luminescence

EnSpire ultra-sensitive luminescence

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- Dedicated PMT for reading luminescence
- PMT is positioned as close as possible over the well to read luminescence signal.



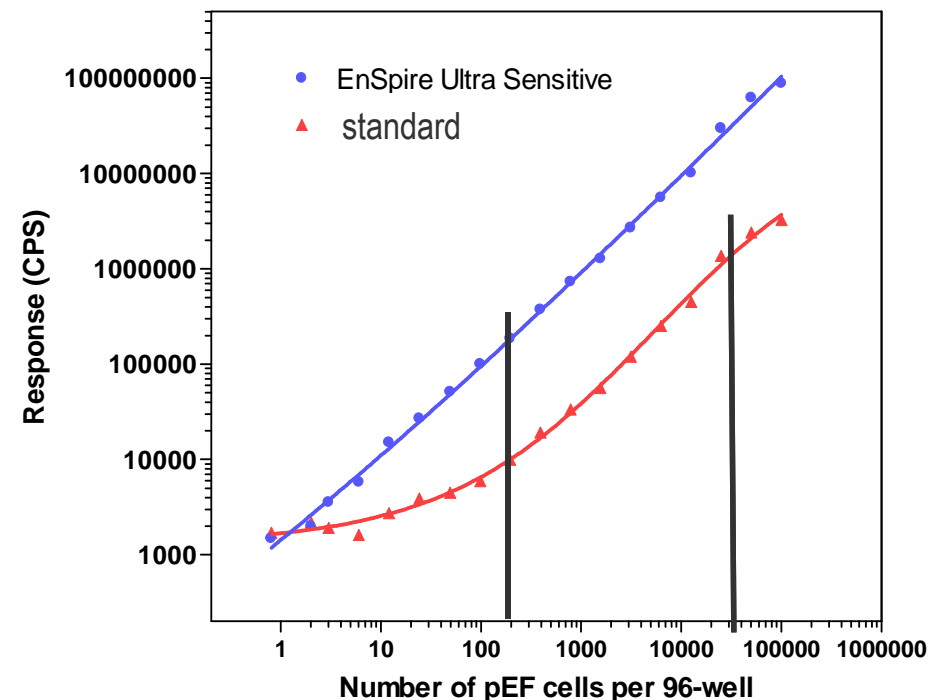
Comparing standard to Ultra Sensitive Luminescence

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- Performance comparison between Standard and Ultra-sensitive luminescence
- Luminescence assay
 - ATPlite cell viability assay
 - Cell numbers ranging from 1 to 100000 cells per well

• Results

- Ultra-sensitive luminescence shows 40 x lower detection limit
- Measurement is linear over whole range of the assay in ultra-sensitive mode whereas it shows only limited linearity in standard mode



EnSpire with dispenser

Dispenser Unit:

- ✓ 2 syringes
- ✓ Waste pump
- ✓ Liquid Temperature control system
- ✓ Magnetic Stirrer



- ✓ The dispenser unit is equipped with two pumps
- ✓ Integrated hot plate (from ambient + 4 °C to 65 °C) and magnetic stirrer (100 - 500 rpm)
- ✓ Dispensing up to 384-well plates
- ✓ Volumes between 1µL .. 475µL
- ✓ Dispense increments 0.5 µl steps

EnSpire Software

Navigation panel – Your Main Dashboard

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- **Always on top**
- **4 Main Functions to Operate**

1. Run Assay
2. Create Protocol
3. Edit Protocol
4. Show Results


Push button to select
meny/execute command


Command Information Bar


- **One button push to select any command menu**
 - Selected command executed by pressing the same button
- **Settings include all system parameters**
- **Next step guidance on command information bar**


Navigation panel – Run Protocol


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Run ABS @ 405


Create Protocols


Edit Protocols


Show Results


Unload Plate


Settings

Protocol name:
ABS @ 405**Plate:**
96 General

Protocol is locked

Navigation panel – Create Protocols

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Run Protocols Create Protocols Edit Protocols Show Results Unload Plate Settings

Please select an option for creating a new protocol

Copy selected or Create new

Valid	Name
	ABS @ 405
	ABS @ 405 On-the-Fly
	Wallac ABS 405 Test Plate
	Wallac ABS 340 Test Plate
	Wallac ABS 450 Test Plate
	Wallac ABS 492 Test Plate
	AlphaScreen 96 Corning Half Area Plate
	AlphaScreen TNFalpha Receptor Binding kit
	AlphaScreen TruHits kit

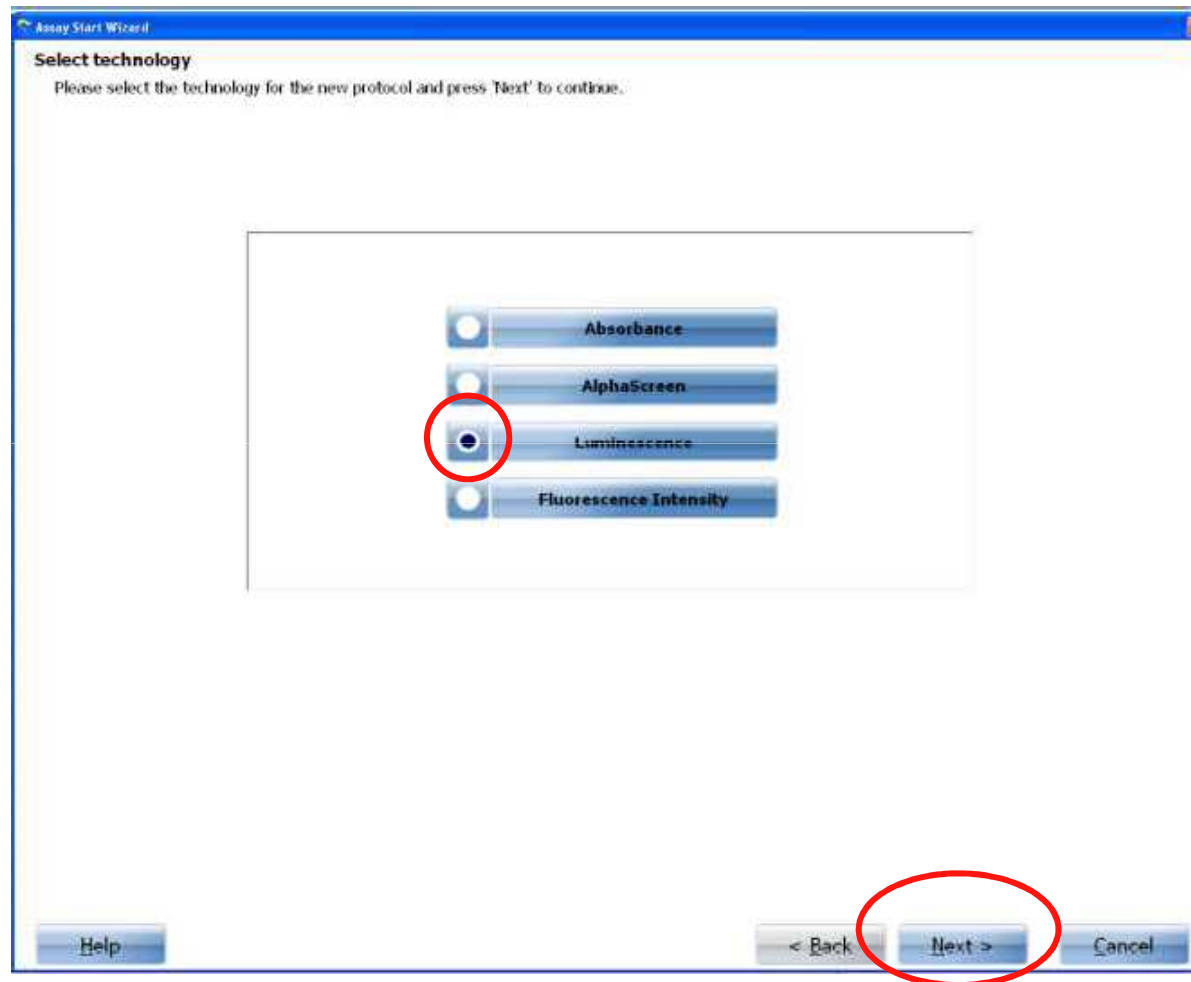
Protocol name:
ABS @ 405

Plate:
96 General

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

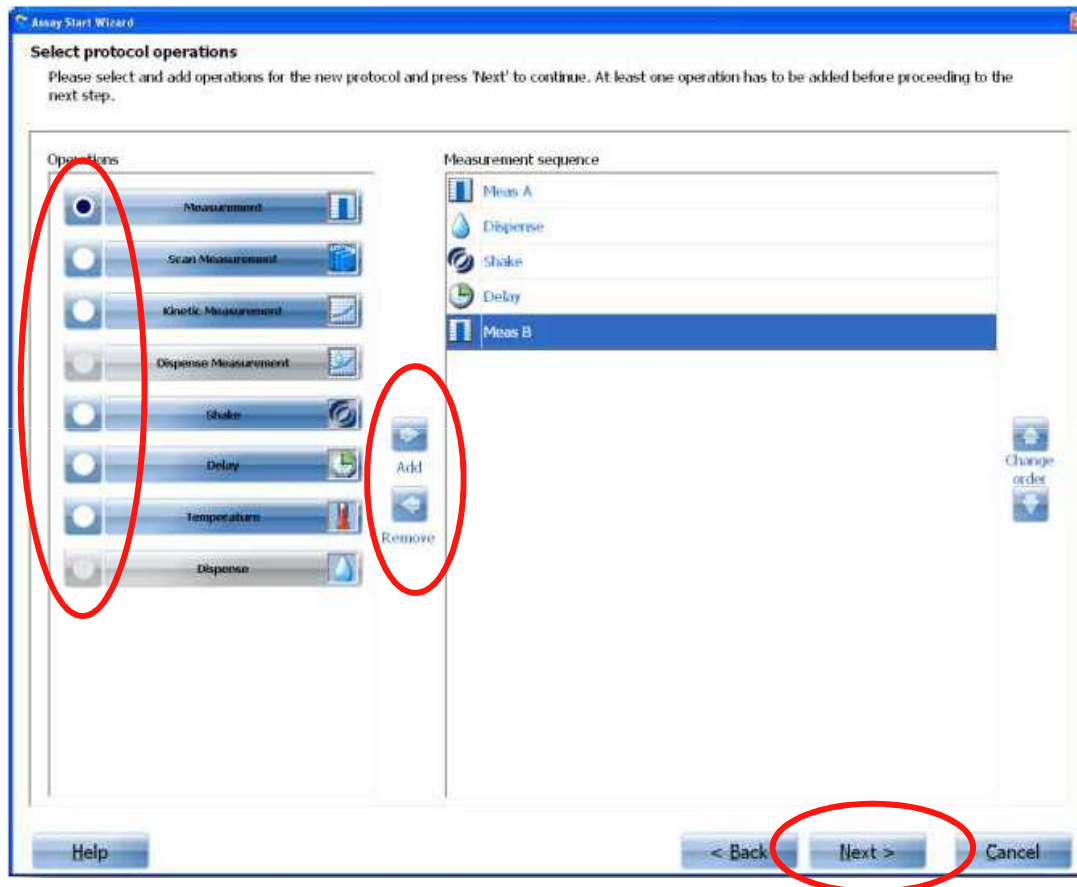
Navigation panel – Create Protocols

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Navigation panel – Create Protocols

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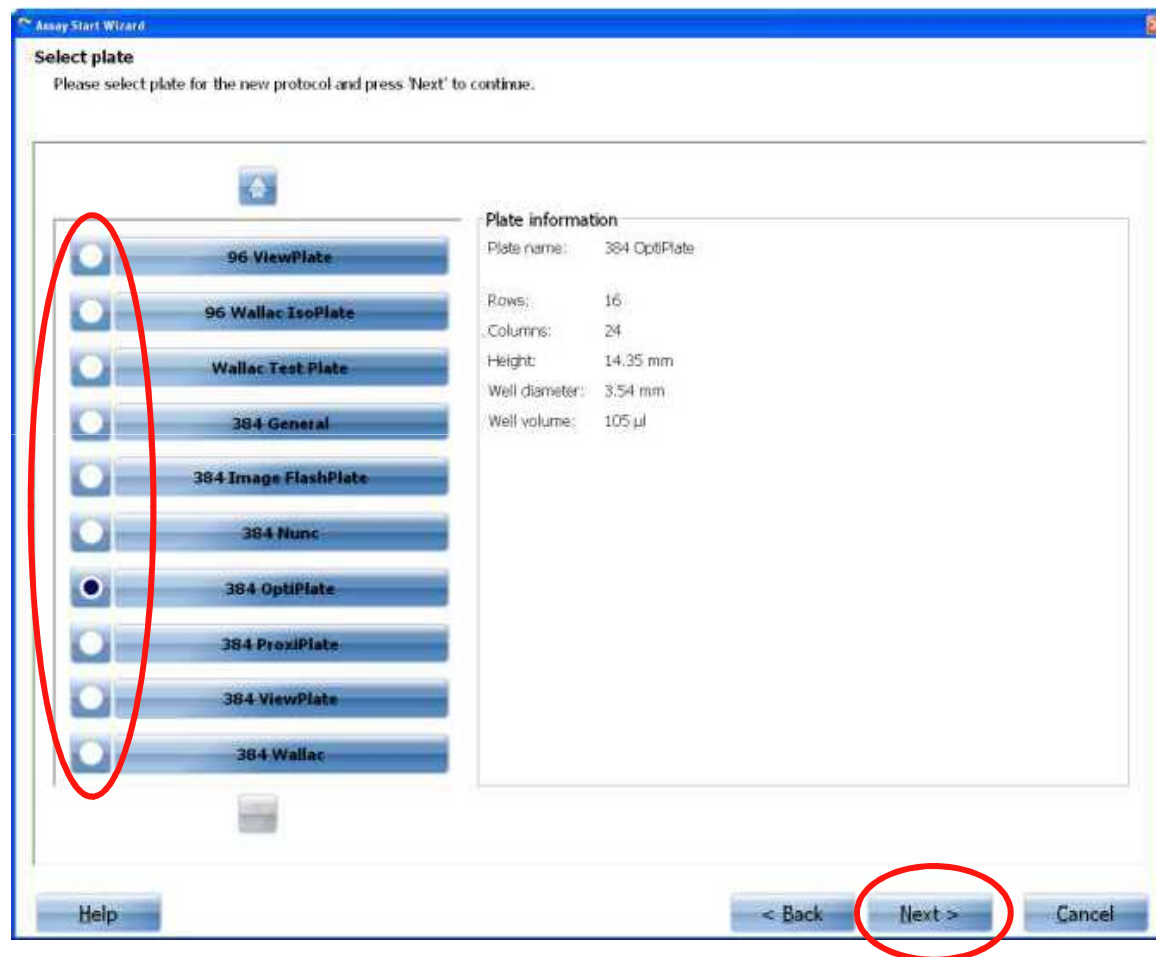


- ➔ 從偵測方式中選一個：
 - Measurement – 一般偵測
 - Scan Measurement – 多點偵測
 - Kinetic Measurement – 連續偵測
 - On-the-fly Measurement – 快速模式
 - Wavelength Scan Measurement – 掃描樣品吸收光譜功能
- ➔ 再選擇是否
 - Shake – 震盪微量盤
 - Delay – 等待時間
- ➔ 選擇時按 Add 將選項放入 Measurement sequence 內，取消時使用 Remove 移除。
先加入動作，再將不需要的移除！
- ➔ Measurement sequence 內順序
以上下箭頭變動。

Navigation panel – Create Protocols

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Choose plate type

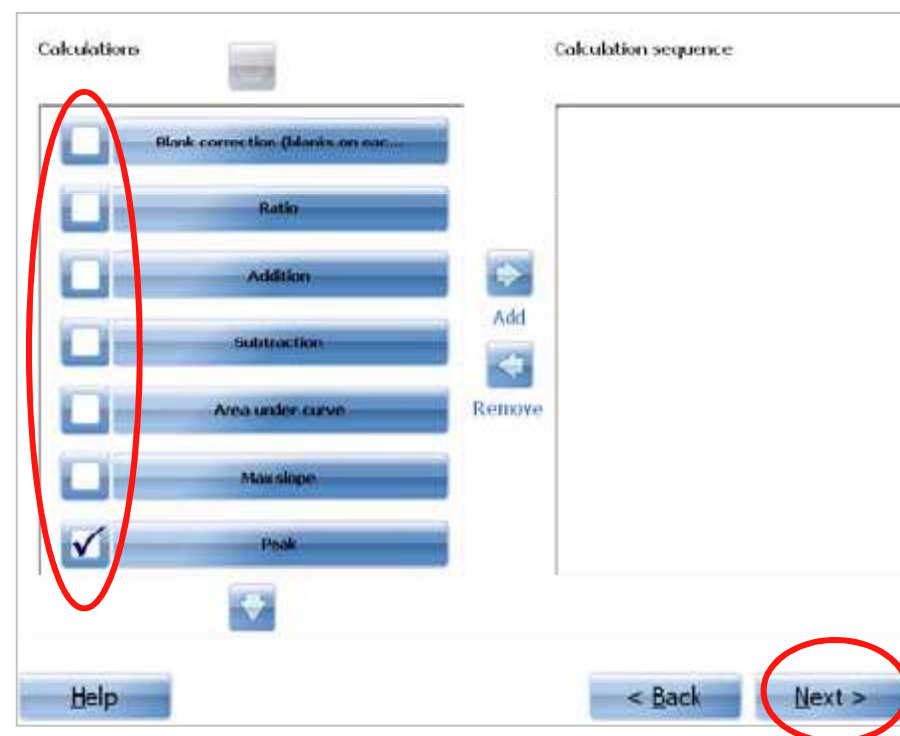
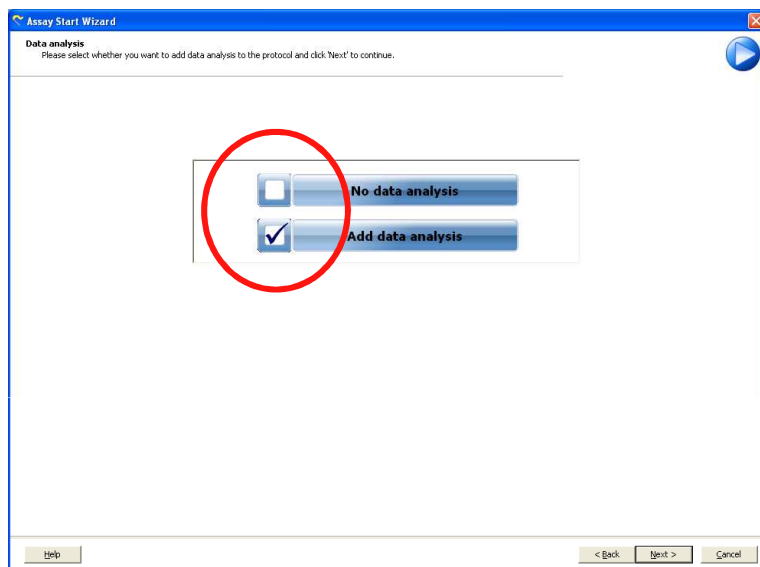


* add new plate type :
Settings - Inventory - Plates

Navigation panel – Create Protocols

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Data analysis



Navigation panel – Create Protocols

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Select well options

Assay Start Wizard

Select well options
Please select options for plate wells and click 'Next' to continue.

Plate 96 Corning Half Area Plate

	1	2	3	4	5	6	7	8	9	10	11	12
A	BL	BL	BL	BL	BL	BL	-	-	-	-	-	-
B	-	-	-	-	-	-	-	-	-	-	-	-
C	STD1	STD2	STD3	STD4	STD5	STD6	STD7	STD8	STD9	-	-	-
D	-	-	-	-	-	-	-	-	-	-	-	-
E	-	-	-	-	-	-	-	-	-	-	-	-
F	-	-	-	-	-	-	-	-	-	-	-	-
G	-	-	-	-	-	-	-	-	-	-	-	-
H	-	-	-	-	-	-	-	-	-	-	-	-

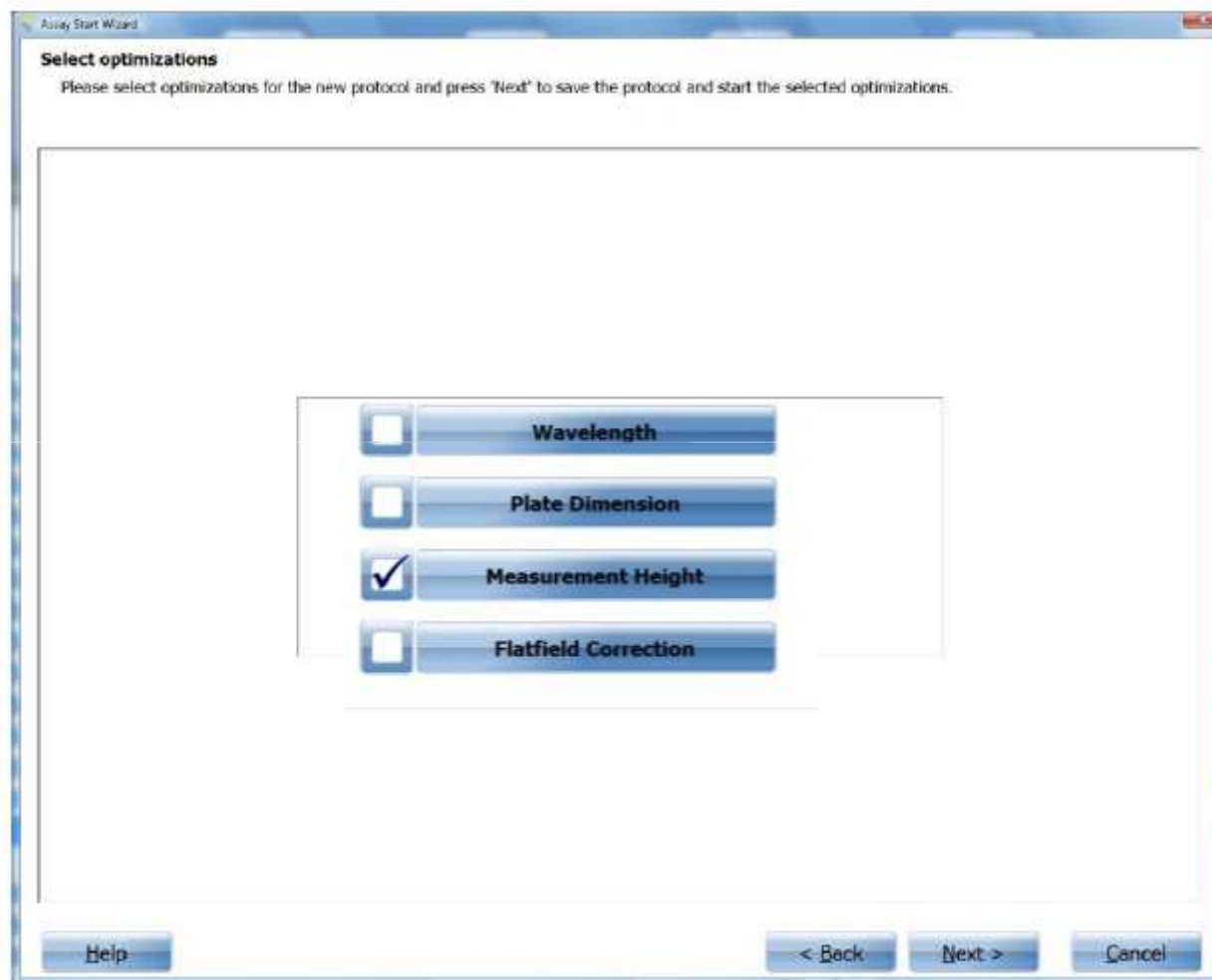
Well options

- ☐ Delete selected
- ☐ LANCE_Blank
- ☐ LANCE_Crosstalk
- ☐ LANCE_High
- ☒ Blank
- ☐ Undefined
- ☐ Control
- ☐ Standard
- ☐ Unknown
- ☐ PL Sample
- ☐ Z_Low
- ☐ Z_High

Replicates

Help < Back Next > Cancel

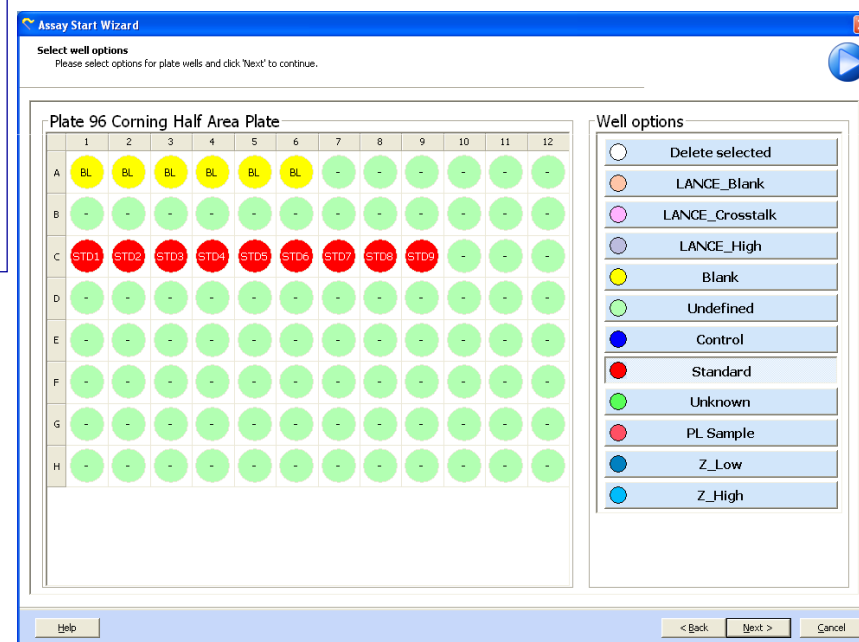
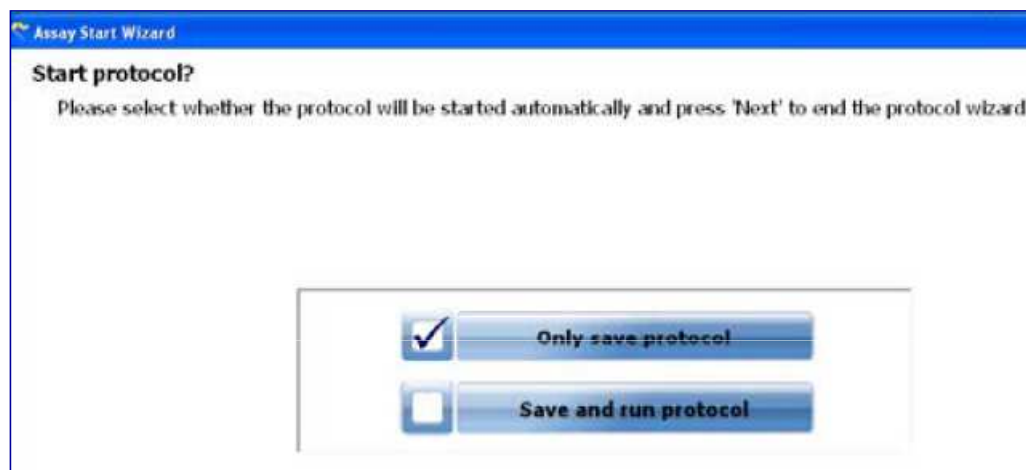
Select optimizations



Navigation panel – Create Protocols

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Save Optimization



Navigation panel –Edit Protocols

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Run Protocols Create Protocols **Edit Copy of Luminescence 384** Show Results Load Plans Settings

Please select a protocol and press the 'Edit' button above

Temperature control not activated. Chamber 22.6 °C Idle (Simulation mode) Help

Valid	Technology	Name
<input checked="" type="checkbox"/>	Lum	Copy of Luminescence 384
<input checked="" type="checkbox"/>	Abs	Copy of Fluorescein
<input checked="" type="checkbox"/>	Abs	ghnfhghfgj
<input checked="" type="checkbox"/>	Abs	Abs
<input checked="" type="checkbox"/>	FI	New
<input checked="" type="checkbox"/>	Abs	Test
<input checked="" type="checkbox"/>	Lum	TEst
<input checked="" type="checkbox"/>	FI	Test
<input checked="" type="checkbox"/>	FI	Test
<input checked="" type="checkbox"/>	Lum	Test
<input checked="" type="checkbox"/>	FI	Test
<input checked="" type="checkbox"/>	Alpha	AS Test
<input checked="" type="checkbox"/>	Lum	Lum Test
<input checked="" type="checkbox"/>	Abs	ghfgh
<input type="checkbox"/>	Lum	LuminTest

Protocol name:
Copy of Luminescence 384

Measurement technology:
Lum

Plate:
384 General

Multiplate assay: No

Export protocol

Delete protocol

Navigation panel – Results

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The screenshot displays the PerkinElmer software interface. At the top, there is a navigation bar with five icons: Run Protocols, Create Protocols, Edit Protocols, Show Wallac ABS 405 Test Plate (highlighted with a yellow border), and Unload Plate. Below the navigation bar, a message states: "Please select a result and press the 'Show' button above". The main area is divided into two panels. The left panel, titled "Assay : Protocol", shows a list with "1 Wallac ABS 405 Test Plate" selected. The right panel displays details for the selected protocol: "Protocol name: Wallac ABS 405 Test Plate", "Plate: Wallac Test Plate", and "Finished: 12/12/2008 10:21:26 AM". Below this information is a 12x12 grid representing a test plate. A red dot is visible in the first column of the grid. At the bottom of the right panel is a "Show results" button.

Run Protocols Create Protocols Edit Protocols Show Wallac ABS 405 Test Plate Unload Plate Settings

Please select a result and press the 'Show' button above

Assay : Protocol

1 Wallac ABS 405 Test Plate

Protocol name:
Wallac ABS 405 Test Plate

Plate:
Wallac Test Plate

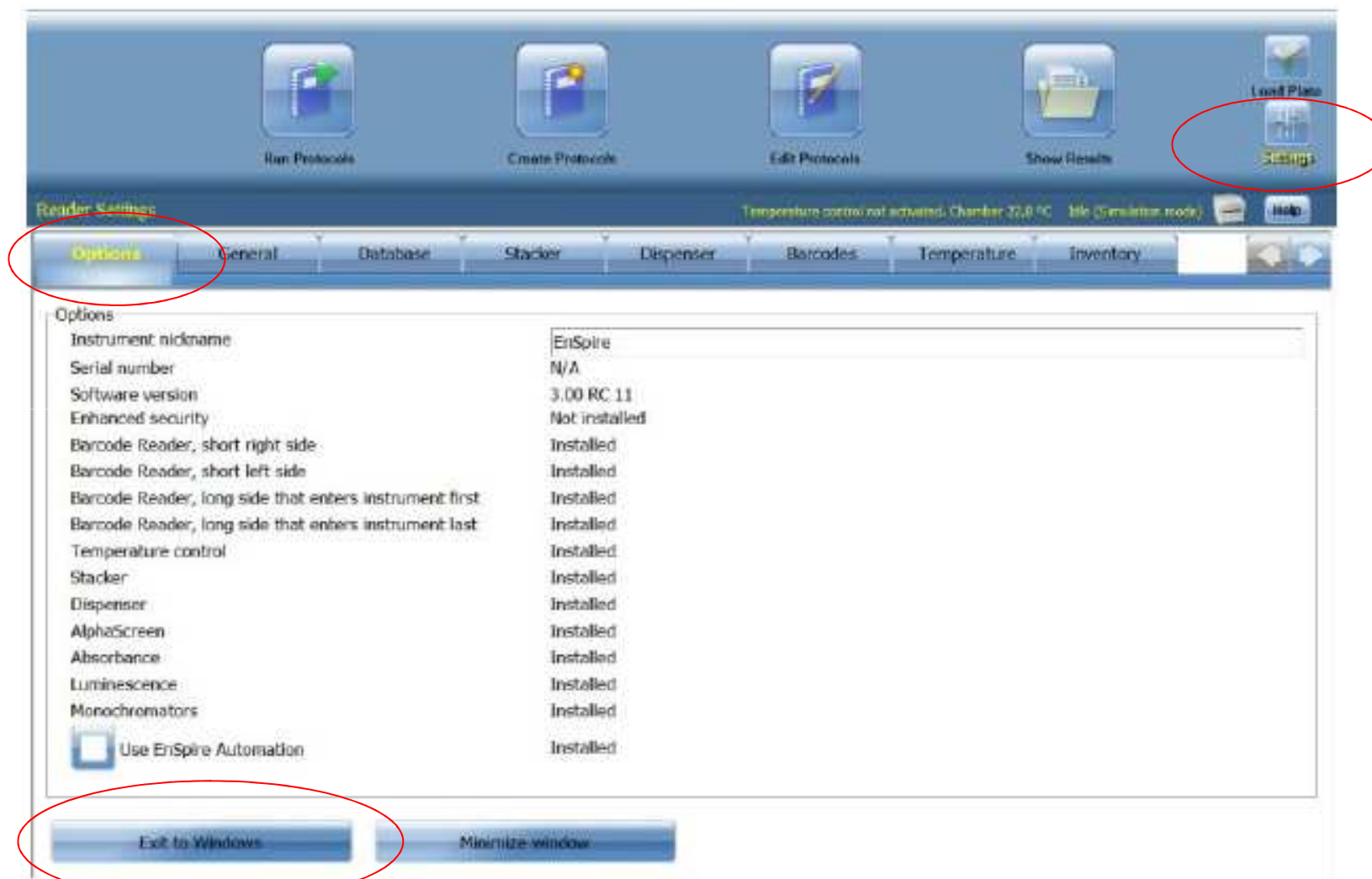
Finished:
12/12/2008 10:21:26 AM

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

Show results

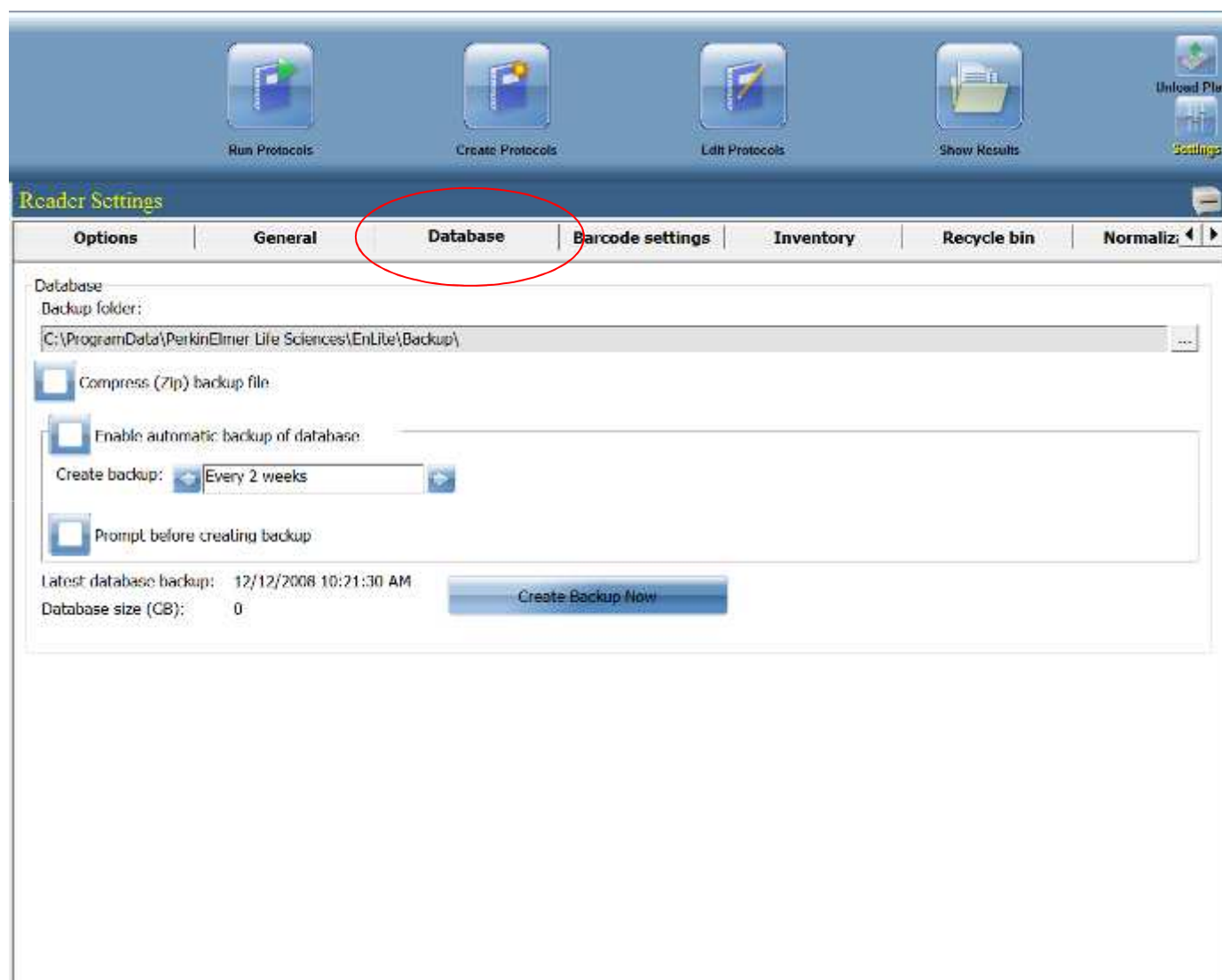
Navigatin panel – Settings...options

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Settings...database

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Settings...barcode settings

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The screenshot displays the 'Reader Settings' window in the PerkinElmer software. The 'Barcode settings' tab is selected and highlighted with a red circle. The interface includes a top toolbar with icons for 'Run Protocols', 'Create Protocols', 'Edit Protocols', 'Show Results', 'Unload Plate', and 'Settings'. The 'Barcode settings' tab contains several sections: 'Barcode reading' with sub-tabs 'Barcode reading' and 'Protocol starting'; 'Read barcode from the' with four options (short right side, short left side, long side first, long side last) each with a checkbox and a 'barcode' button; 'Protocol definition by barcodes' with options to use barcodes as plate ID, define protocol, define plate ID, and split barcodes; and 'Plates without ID barcodes' with options to replace missing plate ID and time stamp.

Reader Settings

Options | **General** | **Database** | **Barcode settings** | **Inventory** | **Recycle bin** | **Normaliz**

Barcode reading | Protocol starting

Read barcode from the

- ☐ short right side and use it as [barcode]
- ☐ short left side and use it as [barcode]
- ☐ long side that enters instrument first and use it as [barcode]
- ☐ long side that enters instrument last and use it as [barcode]

Protocol definition by barcodes

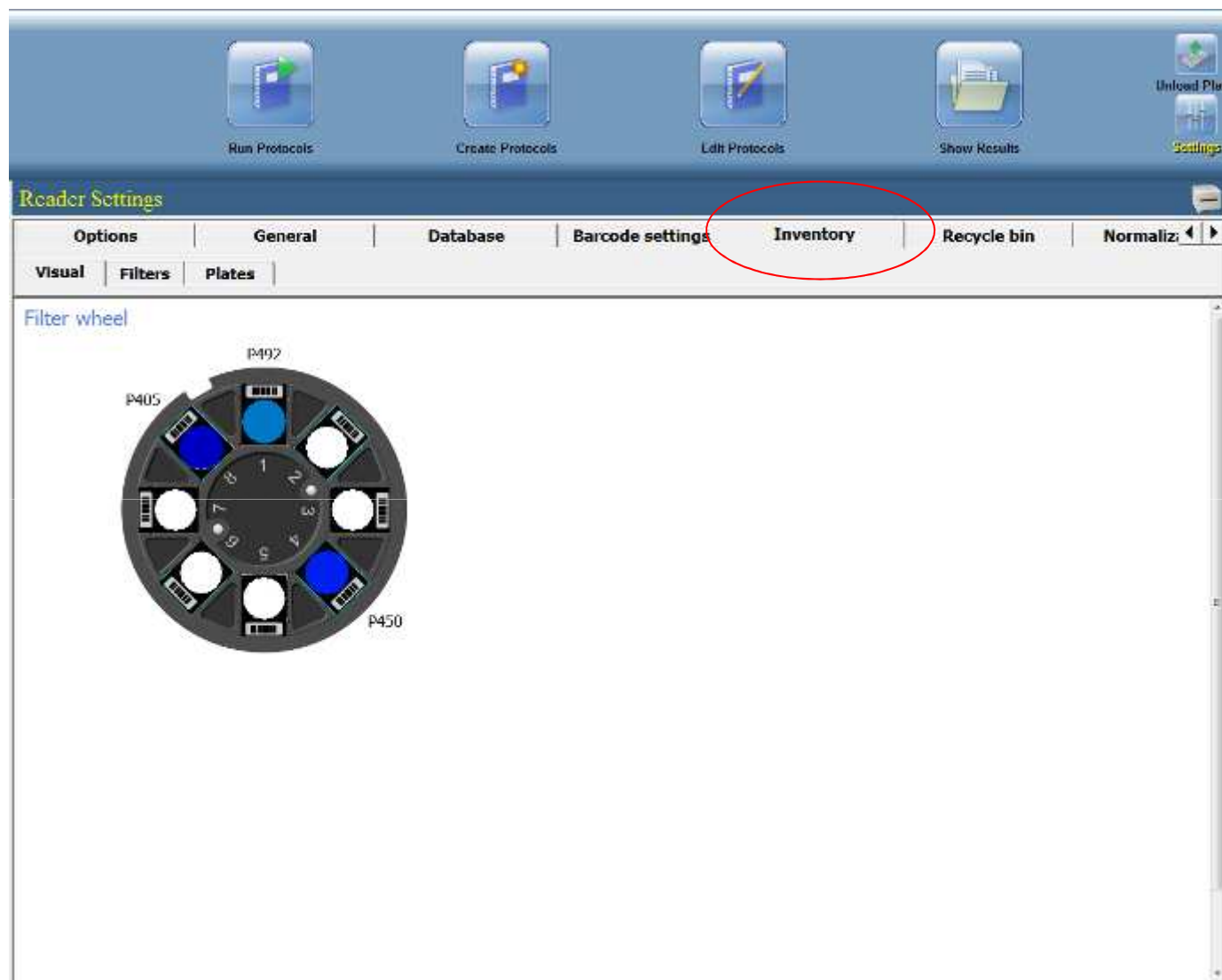
- ☒ Use barcodes as plate ID only
- ☐ Define the protocol using [barcode]
- ☐ Define the plate ID using [barcode]
- ☐ Split barcode:
 - ☐ First [5] digits define [protocol] [barcode]
 - ☐ None of [] digits defines plate ID barcode

Plates without ID barcodes

- ☐ Replace missing Plate ID barcode with
- ☒ Time stamp
- ☐ Custom text [] []

Settings...inventory

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THANK YOU